

Novel Chemo-Enzymatic Synthesis of Optically Active Platelet Activating Factor¹

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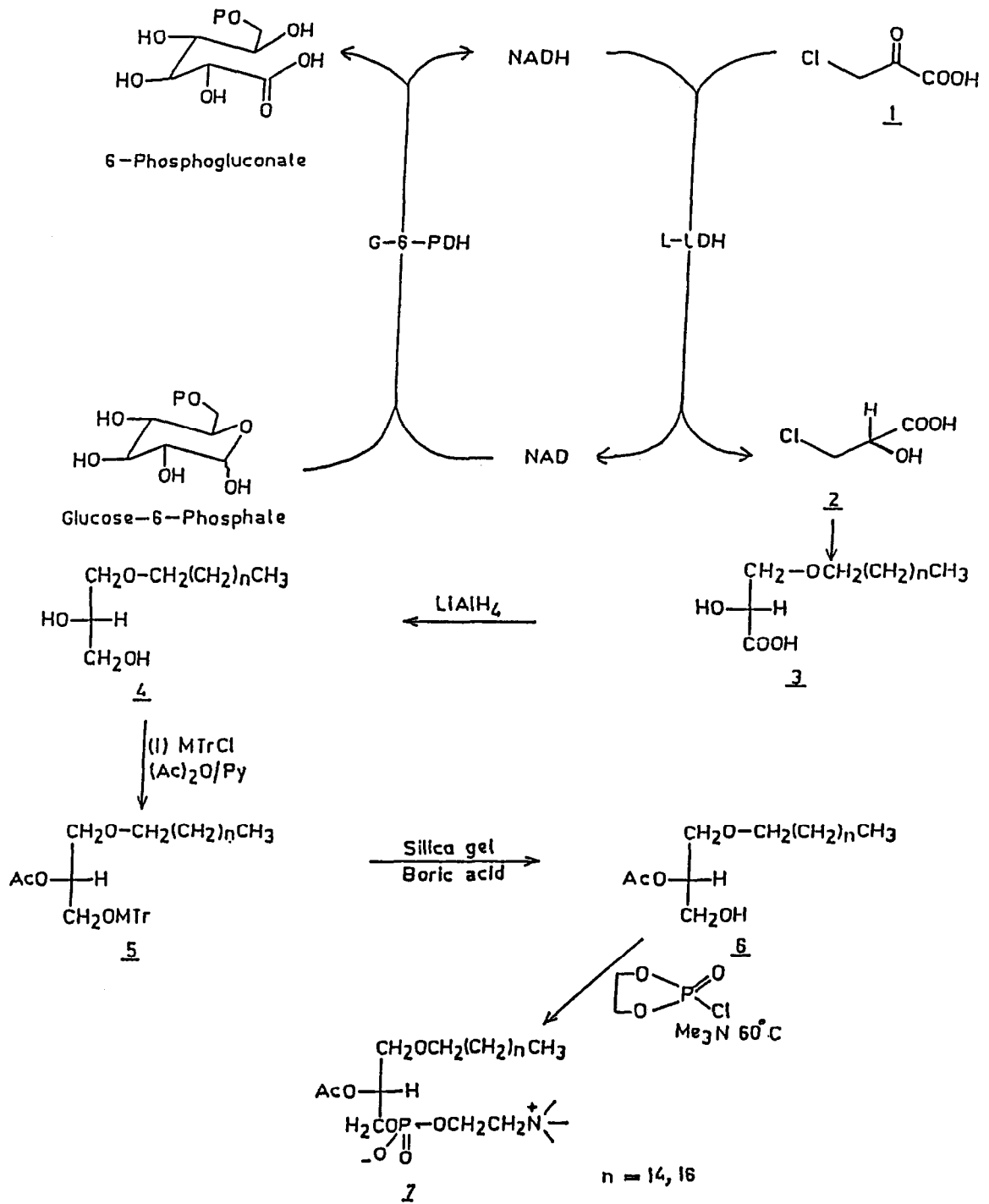
Abstract : This paper describes an enantioselective enzymatic synthesis of biologically active platelet activating factor (PAF) starting from chloropyruvic acid.

Platelet activating factor (PAF), 2-acetyl glyceryl ether phosphorcholines are chemically members of phospholipid family. They have attracted considerable interest principally owing to their pronounced biologically responses²⁻⁵.

Present report describes a straight forward and shortest synthesis of PAF starting from chloropyruvic acid 1. Chloropyruvic acid on L-lactic dehydrogenase catalyzed reduction with NADH yields (L)-chlorolactic acid 2.

The conditions used to synthesize 2 have the following constrains : firstly, the reducing agent in the system NADH is very expensive to be used in stoichiometric ratio. Secondly, the halopyruvic acid deactivates lactic dehydrogenase (presumably by alkylation). In order to overcome these constrains we have used a recycling system (glucose-6-phosphate/glucose-6-phosphate dehydrogenase). The deactivation of lactatedehydrogenase was minimized by using less reactive chlorpyruvic acid. The concentration of chloropyruvic acid was maintained close to K_m by slow addition⁶.

L-chlorolactic acid was synthesized from chloropyruvic acid by the following method : A mixture of dithiothretol (3.6 mM), EDTA (1.4 mM), $MgCl_2$ (7.0 mM) and glucose-6-phosphate (0.51 mol) was taken and its pH adjusted at 7.6 with KOH. The solution was degassed and to it was added NAD (0.53 mmol) and aqueous suspension of (L)-LDH (2300 units) and G-6 PDH (280 units). The pH of the reaction was maintained in between 7.4-8 by addition of 1.0 M KOH solution. An aqueous



solution of chloropyruvic acid (1.0 M, maintained at 5°C) was added dropwise. A total of (0.5 mol) of chloropyruvic acid and (1.27 mmol) of NAD were added over the course of reaction. After 72 hrs the reaction mixture was washed with 0.05 M HEPES buffer (pH 7.5) separated by centrifugation and resuspended in fresh buffer. To this solution BaCl_2 was added followed by addition of 2 ml of ethanol. The precipitate formed (which is L-chlorolactic acid) was filtered, washed and dried⁶ mp 88-89°C $[\alpha]_D +4.14$ (c, 10 g/100 H_2O).

L-chlorolactic acid on reaction with hexadecanol or octadecanol in presence of NaH gives 3⁷ which on reduction with lithium aluminum hydride afforded diol 4. Diol on reaction with 4-methoxy tritylchloride (MTr-Cl) followed by acetylation of the free hydroxy group with acetic anhydride and pyridine gave 5. The selective removal of MTr-group (without migration of acetyl group taking place) was done by passing a solution of 5 in pet ether through a short silica/boric acid column to yield 6. Phosphorylation of 6 with 2-chloro-2-oxo 1,3,2 dioxaphospholane in presence of trimethylamine in benzene⁸ finally yielded the desired L-PAF 7⁹⁻¹².

References and Notes

1. Communication number 5051 from Central Drug Research Institute.
2. Honma, Y.; Kasukabe, M.; Hozumi, M.; Tsaushima, S. and Novmura, H. Cancer Res. 1981, 41, 3211.
3. Wissner, A.; Kholer, C.A. and Goldstein, B.M. J. Med. Chem. 1985, 28, 1365.
4. Cusack, N.J. Nature, 1981, 285, 193.
5. Fujika, K.; Makai, A.; Koboyashi, S.; Inoue, K.; Nujima, S. and Onna, M. Tetrahedron Lett. 1982, 23, 3507.
6. Wong, C.H. and Witeside, G.M. J. Am. Chem. Soc. 1981, 103, 4890.
7. Pollak, A.; Blumenfeld, H.; Baughn, R.L. and Whitesides, G.M. J. Am. Chem. Soc. 1980, 102, 6324.
8. Chandrakumar, N.S. and Hajudu, J. Tetrahedron Lett. 1982, 23, 1043.
9. Wissner, A. J. Org. Chem. 1979, 44, 4617.
10. Kumar, A. and Shanker, K. Synthetic Commun. 1991, 22, 1763.
11. M.S.: n=14 M^+ 532; n=16 M^+ 567; m.p. : n=14, 248°C; n=16, 265°C; $[\alpha]_D$ n=14 +3.50 (methanol); $[\alpha]_D$ n=16 +2.35 (methanol); IR (KBr) 1730, 1240, 1210 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3) 0.84 (t, 3H, (CH_3)), 1.20-1.50 (m 28H, (CH_2)_n), 3.46 (t, 2H, O- CH_2), 3.59 (d, 2H, CH_2 -O-CO) 2.05 (s, 3H, O-CO- CH_3), 5.18 (q, 1H, CH -oAc) 4.01 (m, 2H, CH_2 -O-P), 4.30 (bm, 2H, P-O- CH_2), 3.75 (t, 2H, CH_2 - N^+ -

$(\text{CH}_3)_3$, 3.36 (s, 9H, $-\overset{+}{\text{N}}(\text{CH}_3)_3$).

12. Heymans, F.; Michel, E.; Borrel, M.C.; Wichrowski, B.; Godfroid, J.J.; Convert, O; Coeffier, E.; Tence, M; Benveniste, J. Biochim. Biophys. Acta 1981, 666, 230.

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